HOOK™ GST Protein Spin Purification (Yeast)

For the Purification of GST-Tagged Proteins from Yeast

(Cat. # 786-642)
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INTRODUCTION
HOOK™ GST Protein Spin Purification kit allows for the rapid purification of soluble, GST tagged protein from yeast cultures. The yeast are first lysed with Yeast PE LB™ and LongLife™ Zymolyase® to release total soluble protein, whilst maintaining the structure and activity of the protein. The GST tagged protein is purified by affinity chromatography by adding 0.5ml Glutathione resin to the clarified lysate. The resin is transferred to a convenient spin column, where it is rapidly washed and the GST protein is eluted.

Yeast-PE LB™ is useful for extraction of soluble proteins from yeast cells. Yeast PE LB™ is a proprietary improvement on the Zymolyase® based spheroplast preparation and extraction of soluble proteins from yeast cells. Yeast PE LB™ is based on organic buffering agents that utilize a mild non-ionic detergent and a proprietary combination of various salts and agents to enhance extraction and stability of proteins. A ready-to-use Zymolyase® preparation is also provided. Depending on application, additional agents such as protease inhibitors may be added into Yeast PE LB™. The proprietary combination of this reagent provides a simple and versatile method of yeast protein extraction. Yeast PE LB™ eliminates the need for laborious glass bead lysis of yeast cells. HOOK™ GST Protein Spin Purification kit is optimized to yield ~1mg of soluble GST tagged protein, with a purity of 80-90%, dependent on expression levels, resin type, conformation and solubility characteristics of the protein.

ITEM(S) SUPPLIED (Cat. # 786-642)

<table>
<thead>
<tr>
<th>Description</th>
<th>Size</th>
</tr>
</thead>
<tbody>
<tr>
<td>Yeast Suspension Buffer</td>
<td>15ml</td>
</tr>
<tr>
<td>Yeast PE-LB™</td>
<td>100ml</td>
</tr>
<tr>
<td>LongLife™ Zymolyase®</td>
<td>2 x 0.5ml</td>
</tr>
<tr>
<td>Glutathione Resin*</td>
<td>12.5ml</td>
</tr>
<tr>
<td>GST Binding/Wash Buffer</td>
<td>200ml</td>
</tr>
<tr>
<td>Glutathione</td>
<td>2 vials</td>
</tr>
<tr>
<td>Spin Column</td>
<td>25</td>
</tr>
<tr>
<td>Caps (Micro, screw cap)</td>
<td>25</td>
</tr>
<tr>
<td>Collection Tubes</td>
<td>50</td>
</tr>
</tbody>
</table>

*Glutathione Resin is supplied as 25ml 50% slurry in 20% ethanol.
STORAGE CONDITION
The kit is shipped at ambient temperature. Upon arrival, store LongLife™ Zymolyase® at -20°C, resin refrigerated at 4°C (DO NOT FREEZE), and all other components may be stored at room temperature. The kit components are stable for 1 year when stored and used as recommended.

PREPARATION BEFORE USE
1. Prior to using the HOOK™ GST Protein Spin Purification kit, it is recommended that an estimation of the expression and solubility levels of your protein is performed. Express protein as normal and lyse with the Yeast PE-LB™ reagents, clarify by centrifugation and view on a SDS polyacrylamide gel.
2. To maintain the integrity of your recombinant protein, it is recommended that a protease inhibitor cocktail is used throughout the purification process. We recommend Yeast & Fungal ProteaseARREST™ (Cat. # 786-333), a protease inhibitor cocktail specific designed for purifying proteins from yeast, or ProteaseARREST™ (Cat. # 786-108), a general protease inhibitor that is supplied with optional EDTA.
3. The resin and buffers should be allowed to equilibrate to room temperature before beginning the purification.
4. GST Elution Buffer: Dissolve one vial of glutathione in 50ml GST Binding/Wash Buffer to give a final concentration of 10mM. For long term storage, store at -20°C. We recommend freezing in smaller aliquots to limit the amount of freeze/thaws and potential oxidation of the glutathione.

ADDITIONAL ITEM(S) REQUIRED
• Centrifuge and centrifuge tubes for harvesting yeast cultures
• Wide-bore pipette tips for dispensing the resin slurry
• Micro-centrifuge
• 15ml centrifuge tubes
PROTOCOL

1. Harvest the yeast cells from an overnight culture (OD$_{600}$ 1.5-2.0) by centrifugation at 5,000xg for 10 minutes. Discard the supernatant.

   **NOTE:** If using a frozen cell pellet, ensure the pellet is completely thawed before starting.

2. Resuspend a maximum of 0.5ml cell pellet in 0.5ml Yeast Suspension Buffer.

   **NOTE:** If using, add your protease inhibitor cocktail to the suspension at this point. For Yeast & Fungal ProteaseARREST™ (Cat. # 786-333) or ProteaseARREST™ (Cat. # 786-108), add 45µl.

3. Vortex for 1 minute or until the cell suspension is homogeneous. Incubate the suspension for 5 minutes at 4°C. Vortex again to suspend the cells.

4. Briefly, vortex the LongLife™ Zymolyase® to mix the solution. Add 40µl LongLife™ Zymolyase® to the cell suspension. Gently mix the content.

5. Incubate the suspension at 37°C for 30-60 minutes.

6. At the end of incubation, centrifuge the suspension at 10,000x g for 5 minutes. Remove and discard the supernatant carefully, leaving the spheroplast pellet in the tube.

7. Suspend the spheroplast pellet in 4ml Yeast-PE LB™ Buffer. Pipette the suspension up and down a few times. Vortex periodically and incubate on ice for 30 minutes. Incubating the cells for 1-3 minutes at 37°C or a brief sonication step may further facilitate the lysis.

8. Centrifuge at 20,000x g for 30 minutes at 4°C. Transfer the clarified lysate to a 15ml centrifuge tube.

9. Swirl the resin bottle to achieve a homogenous slurry and, using a wide bore pipette tip, transfer 1ml 50% resin slurry to the yeast lysate. Close the tube.

10. Incubate, with shaking or rotation, for 15 minutes at room temperature.

11. Centrifuge the tube at 1,500xg for 5 minutes to pellet the resin.

12. Discard the supernatant, without disturbing the resin, and then resuspend the resin in 250µl Wash Buffer.

13. Snap off the tab on the base of the spin column and place in a collection tube. Transfer the resin suspension to the spin column using a wide bore pipette tip. Centrifuge the spin column assembly at 2,000xg for 2 minutes. Discard the flow through and return the spin column to the collection tube.

14. Add 500µl Wash Buffer to the spin column to wash away unbound and non-specific proteins. Incubate at room temperature for 5 minutes and then centrifuge at 2,000xg for 2 minutes. Discard the flow through and return the spin column to the collection tube.

   **NOTE:** If there is an issue with a large amount of non-specific proteins binding then the amount of low level of competing imidazole in the wash buffer may be increased.

15. Repeat step 11 once. Transfer the spin column to a clean collection tube.
16. Elute the GST tagged protein by adding 0.5ml Elution buffer to the resin and incubating at room temperature for 5 minutes. Centrifuge the spin column assembly at 2,000xg for 2 minutes. Transfer the flow through to a 1.5ml centrifuge tube and return the spin column to the collection tube. Repeat the elution three more times, storing each elution in a different 1.5ml centrifuge tube.

17. The elution of the protein can be monitored by absorption at 280nm, by assaying with a protein assay (CB-X™ Protein Assay (Cat. # 786-12X) or by SDS-PAGE analysis. We recommend Tube-O-DIALYZER™ for buffer exchange and removal of excess imidazole.
## TROUBLESHOOTING

<table>
<thead>
<tr>
<th>Issue</th>
<th>Possible Cause</th>
<th>Suggested Solution</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Low Protein Yield</strong></td>
<td>Poor expression of soluble protein</td>
<td>Optimize yeast expression and growth conditions. Check expression by SDS-PAGE to confirm expression.</td>
</tr>
<tr>
<td></td>
<td>The GST tag may not bind column</td>
<td>Check the sequence of the construct to ensure the tag is in frame with the protein of interest. Test for presence of the GST tag by Western blotting and probing with a α-His antibody</td>
</tr>
<tr>
<td><strong>Protein Degradation</strong></td>
<td>Protein is degraded by bacterial proteases</td>
<td>Use a protease inhibitor cocktail that does not use metal chelators. We recommend <em>Yeast/Fungal ProteaseARREST™</em> (Cat. # 786-333), a protease inhibitor cocktail specific designed for purifying proteins from yeast.</td>
</tr>
<tr>
<td><strong>Poor Protein Purity</strong></td>
<td>Poor column washing</td>
<td>Wash the column more than twice or try increasing the imidazole concentration.</td>
</tr>
<tr>
<td><strong>Slow Column Flow</strong></td>
<td>Column overloaded or particulates added to column</td>
<td>Ensure the bacterial lysate is completely clear before adding resin, if necessary centrifuge the lysate a second time</td>
</tr>
</tbody>
</table>

## RELATED PRODUCTS

Download our Protein Cross-Linkers Handbook.

For other related products, visit our website at [www.GBiosciences.com](http://www.GBiosciences.com) or contact us.

Last saved: 06/03/2014 AB